0957-4166(95)00248-0

## Enzymatic Resolution of $\alpha$ -Acetoxysulfides: A New Approach to the Synthesis of Homochiral S,O-Acetals.

John Miltona, Stephen Branda, Martin F. Jonesb and Christopher M. Rayner\*a

<sup>a</sup>School of Chemistry, University of Leeds, Leeds LS2 9JT, U.K.; <sup>b</sup>Chemical Development, Glaxo Research and Development Ltd., Gunnels Wood Road, Stevenage, Herts., SG1 2NY, U.K.

Abstract: A novel enzymatic resolution of  $\alpha$ -acetoxysulfides using *Pseudomonas fluorescens* lipase is reported. Selectivity is highly dependent on the substrate and solvent, with enantiomeric excesses of >95% in some cases. We believe these are the first examples of enzymatic resolutions of an S,O-acetal.

The preparation of homochiral secondary acetates by enzymatic resolution is a well established procedure, often giving very high selectivities with appropriate substrates. Our interest in reactive organosulfur intermediates led us to embark on a programme to develop an efficient synthetic route to Lamivudine (3TC<sup>TM†</sup>, 1), a highly promising drug candidate for HIV<sup>3</sup> and HBV infections. Because of the different toxicities of the two enantiomers of this compound we required a highly enantioselective route and ideally one that would be suitable for large scale synthesis.

Lamivudine 1

The enantioselective synthesis of lamivudine 1 provides a considerable challenge to the synthetic chemist due to the presence of the S,O-acetal and aminal stereogenic centres, both sharing the same oxygen atom.<sup>5</sup> Although a number of synthetic routes to racemic S,O-acetals are available<sup>6</sup>, to date only those derived from 7-thiomenthol originally reported by Eliel<sup>7</sup>, and the asymmetric Pummerer reaction<sup>8</sup>, address the problem of control of absolute stereochemistry, of which only the latter was suitable for use in an approach to lamivudine. Despite the particularly encouraging work recently reported by Kita *et al.*<sup>8a-c</sup>, our systems gave only minimal chirality transfer from the sulfoxide, and this approach was also limited due to problems accessing the required homochiral sulfoxide precursor.<sup>9</sup> We thus needed an alternative strategy for the

1904 J. MILTON et al.

preparation of homochiral  $\alpha$ -acetoxysulfides, which led us to consider enzymatic methods. A literature search failed to reveal any examples of enzymatic resolutions of S,O-stereogenic centres.<sup>1</sup> We thus embarked on a systematic study of this interesting and potentially very useful reaction.

The  $\alpha$ -acetoxysulfide substrates 3 could be prepared by two routes (scheme 1). Coupling of a mercapto acetate<sup>10</sup> 2 with an alkyl halide, followed by oxidation to the sulfoxide and Pummerer rearrangement (route 1) generally worked well. Alternatively, addition of a thiol<sup>10</sup> to methyl glyoxalate<sup>11</sup> and *in situ* acetylation of the intermediate hemiacetal (route 2) provided a shorter, more convenient procedure.

## Route 1

## Route 2

Scheme 1.

With these substrates in hand, we then began to investigate the enzymatic hydrolysis reaction. We initially chose *Pseudomonas fluorescens* lipase because of its reported efficiency for the resolution of more conventional substrates. We were very pleased to observe that the presence of the sulfur atom made no difference to the efficiency of the reaction (table 1). In all cases, exclusive hydrolysis of the acetate group was observed with no detectable hydrolysis of the other ester functionalities. Although the hemithioacetal 5 byproduct could clearly be seen in crude NMR spectra, on purification using column chromatography, only the resolved acetate and thiol (from decomposition of 5) could be isolated. 12

The stereoselectivity of the hydrolysis can be rationalised as being controlled by the different sizes of substitutents either side of the acetate chiral centre. With menthyl as the large group (table 1, entries 1-3), an increase in size of the smaller group reduces the selectivity in the hydrolysis from >95% to <10%. Changing to the methyl ester ( $R^1 = CH_3$ ) allows high selectivity to be observed with large groups in  $R^2$ , with corresponding reversal of absolute stereochemistry at the acetal chiral centre. It would appear, at least in  $CHCl_3$ , that the borderline for this change in selectivity is between the dimethyl and diethyl acetals (entries 4 and 6, note optical rotation). However in BuOMe solvent no reversal in selectivity is observed between these two substrates. Note also that some particularly lipophilic substrates were essentially inert under the reaction conditions in  $CHCl_3$  (entries 8 and 10), but switching to BuOMe as solvent restored reactivity (entries 8 cf. 9; 10 cf. 11), and gave significantly enhanced enantioselectivities (entries 6 cf. 7; 12 cf. 13; 14 cf. 15). Interestingly, in extreme cases, selectivity can begin to decrease if the large group is made too big (entry 9).

In conclusion, we have demonstrated the first enzymatic resolution of an  $\alpha$ -acetoxysulfide achieving very high levels of stereocontrol in many cases, and good yields. We have now used this new chemistry to complete the synthesis of 1 which confirms some of the stereochemical assignments shown in table 1. Further details will be reported shortly. We are currently extending this methodology further to related systems, and developing new synthetic procedures involving the use of homochiral acetals.

Entry	R <sup>1</sup>	P²	Solvent	% Yield <sup>a</sup>	d.e. or e.e. of <b>4</b> (configuration <sup>b</sup> )	$\left[lpha ight]_{D}^{20}$ (c, solvent)
1	Menthyl	-CH₂CH₃	CHCl₃	48	>95° (S)	-26.7 (1.0,EtOH)
2	Menthyl	-CH₂CN	CHCl <sub>3</sub>	47	35 <sup>c</sup> (S)	-36.1 (1.0,EtOH)
3	Menthyl	-CH <sub>2</sub> CH(OCH <sub>3</sub> ) <sub>2</sub>	CHCl3	42	<10 <sup>c</sup>	-48.3 (1.0,EtOH)
4	CH <sub>3</sub>	-CH <sub>2</sub> CH(OCH <sub>3</sub> ) <sub>2</sub>	CHCl₃	49	40 <sup>d</sup> (S)	+13.2 (1.0,EtOH)
5	CH3	-CH <sub>2</sub> CH(OCH <sub>3</sub> ) <sub>2</sub>	<sup>t</sup> BuOMe	45	>95 <sup>d</sup> (R)	-53.5 (3.1,MeOH)
6	CH₃	-CH <sub>2</sub> CH(OEt) <sub>2</sub>	CHCI3	46	30 <sup>d</sup> (R)	-15.6 (1.0,EtOH)
7	CH <sub>3</sub>	-CH <sub>2</sub> CH(OEt) <sub>2</sub>	<sup>t</sup> BuOMe	49	>95 <sup>d</sup> (R)	-31.9 (1.0,EtOH)
8	CH₃	-CH <sub>2</sub> CH(OBn) <sub>2</sub>	CHCl <sub>3</sub>	No reaction		
9	CH₃	-CH <sub>2</sub> CH(OBn) <sub>2</sub>	<sup>t</sup> BuOMe	48	65 <sup>d</sup> (R)	-20.2 (1.0,EtOH)
10	CH₃	-(CH <sub>2</sub> ) <sub>2</sub> CH(OMe) <sub>2</sub>	<sup>t</sup> BuOMe	45	>95 (R)	-9.9 (1.3, EtOH)
11	CH <sub>3</sub>	-¹Bu	СНСІ₃	No reaction		
12	CH₃	- <sup>n</sup> Bu	<sup>t</sup> BuOMe	47	>95 <sup>d</sup> (R)	-41.5 (1.6, CHCl <sub>3</sub> )
13	CH₃	-(CH <sub>2</sub> ) <sub>2</sub> OSiEt <sub>3</sub>	CHCl <sub>3</sub>	42	20 <sup>d</sup> (R)	n.d.
14	CH₃	-(CH <sub>2</sub> ) <sub>2</sub> OSiEt <sub>3</sub>	<sup>t</sup> BuOMe	47	81 <sup>d</sup> (R)	-48.7 (1.7, CHCl <sub>3</sub> )
15	CH₃	-(CH <sub>2</sub> ) <sub>3</sub> OSiEt <sub>3</sub>	CHCl₃	46	25 <sup>d</sup> (R)	n.d.
16	CH <sub>3</sub>	-(CH₂)₃OSiEt₃	<sup>t</sup> BuOMe	48	85 <sup>d</sup> (R)	-36.2 (2.9, CHCl <sub>3</sub> )
17	СН₃	HN OBn	СНСІ₃	43	88° (R)	+20.1 (2.1, CHCl <sub>3</sub> )

<sup>a</sup>Standard conditions: 100mg substrate, phosphate buffer (pH 7, 2ml), solvent (0.5ml), PFL (2mg), 30°C, 2h; reaction had generally proceeded to 50% (±5%) as determined by <sup>1</sup>H NMR; <sup>b</sup>tentatively assigned using literature model<sup>13</sup>; <sup>c</sup>Determined by <sup>1</sup>H NMR; <sup>d</sup>Determined by <sup>1</sup>H NMR using (+)-Eu(hfc)<sub>3</sub> or (-)-2,2,2-trifluoro-1-(9-anthryl)ethanol.

Table 1. Results of hydrolysis experiments.

1906 J. MILTON et al.

Acknowledgements: we thank Glaxo Research and Development for a postdoctoral fellowship (JM), and Glaxo Research and Development and the EPSRC for a CASE award (SB).

†3-TC<sup>™</sup> is a registered trade mark of the Glaxo group of companies.

## References.

- 1. a) Roberts, S.M.; Wiggins, K.; Casy, G.; Phythian, S. Preparative Biotransformations, J. Wiley, Chichester, 1992; b) Davies, H.G.; Green, R.H.; Kelly, D.R.; Roberts, S.M. Biotransformations in Preparative Organic Chemistry, Best Synthetic Methods Series, Academic Press: London, 1992. There is a single report of the enzymatic hydrolysis of chloral acetyl methyl acetal: Chenevert, R.; Desjardins, M.; Gagnon, R. Chemistry Lett., 1990, 33; α-Hydroxyphosphonates have also been resolved using a related procedure: Drescher, M.; Li, Y.-F.; Hammerschmidt, F. Tetrahedron, 1995, 51, 4933.
- 2. Westwell, A.D.; Thornton-Pett, M.; Rayner, C.M. J. Chem. Soc., Perkin Transactions 1, 1995, 847; Pickersgill, I.F.; Marchington, A.P.; Thornton-Pett, M.; Rayner, C.M. J. Chem. Soc., Chem. Commun., 1995, 647; Archer, N.J.; Rayner, C.M.; Bell, D.; Miller, D. Synlett, 1994, 617; Gill, D.M.; Pegg, N.A.; Rayner, C.M. J. Chem. Soc., Perkin Transactions 1, 1993, 1371.
- 3. Coates, J.A.V.; Cammack, N.; Jenkinson, H.J.; Mutton, I.M.; Pearson, B.A.; Storer, R.; Cameron, J.; Penn, C.R. Antimicrob. Agents and Chemotherap., 1992, 36, 202; Schinazi, R.F.; Chu, C.K.; Peck, A.; McMillan, A.; Mathis, R.; Cannon, D.; Jeong, L.-S.; Beach, J.W.; Choi, W.-B.; Yeola, S.; Liotta, D.C. Antimicrob. Agents and Chemotherap., 1992, 672.
- 4. Doong, S.-L.; Tsai, C.-H.; Schinazi, R.F.; Liotta, D.C.; Cheng, Y.-C. Proc. Natl. Acad. Sci. USA, 1992, 88, 8495.
- 5. For alternative approaches to 3TC and related compounds see: Humber, D.C.; Jones, M.F.; Payne, J.J.; Ramsay, M.V.J.; Zacharie, B.; Jin, H.; Siddiqui, A.; Evans, C.A.; Tse, H.L.A.; Mansour, T.S. Tetrahedron Lett., 1992, 33, 4625; Hoong, L.K.; Strange, L.E.; Liotta, D.C.; Koszalka, G.W.; Burns, C.L.; Schinazi, R.F. J. Org. Chem., 1992, 57, 5563; Beach, J.W.; Jeong, L.S.; Alves, A.J.; Pohl, D.; Kim, H.O.; Chang, C.-N.; Doong, S.-L.; Schinazi, R.F.; Cheng, Y.-C.; Chu, C.K. J. Org. Chem., 1992, 57, 2217; Jeong, L.S.; Alves, A.J.; Carrigan, S.W.; Kim, H.O.; Beach, J.W.; Chu, C.K. Tetrahedron Lett., 1992, 33, 595; Choi, W.-B.; Wilson, L.J.; Yeola, S.; Liotta, D.C.; Schinazi, R.F. J. Amer. Chem. Soc., 1991, 113, 9377; Chu, C.K.; Beach, J.W.; Jeong, L.S.; Choi, B.G.; Comer, F.I.; Alves, A.J.; Schinazi, R.F. J. Org. Chem., 1991, 56, 6503. See also: Wang, W.; Jin, H.; Mansour, T.S. Tetrahedron Lett., 1994, 35, 4739; Belleau, B.; Brasili, L.; Chan, L.; DiMarco, M.P.; Zacharie, B.; Nguyen-Ba, N.; Jenkinson, H.J.; Coates, J.A.V.; Cameron, J.M. Bioorg. Med. Chem. Lett., 1993, 3, 1723.
- 6. De Voss, J.J.; Sui, Z. Tetrahedron Lett., 1994, 35, 49; Kim, S.; Park, J.H.; Lee, J.M. Tetrahedron Lett., 1993, 34, 5769; Kusche, A.; Hofmann, R.; Munster, I.; Keiner, P.; Bruckner, R. Tetrahedron Lett., 1991, 32, 467; Masaki, Y.; Serizawa, Y.; Kaki, K. Chemistry Lett., 1985, 1933; Sato, T.; Kobayashi, T.; Gojo, T.; Yoshida, E.; Otera, J.; Nozaki, H.; Chemistry Lett., 1987, 1661; Guindon, Y.; Bernstein, M.A.; Anderson, P.C.; Tetrahedron Lett., 1987, 28, 2225; Kim, S.; Park, J.H.; Lee, S. Tetrahedron Lett., 1989, 30, 6697; Otera, J. Synthesis, 1988, 95; Cohen, T.; Bhupathy, M.; Acc. Chem. Res., 1989, 22, 152.
- Otera, J. Synthesis, 1988, 95; Cohen, T.; Bhupathy, M.; Acc. Chem. Res., 1989, 22, 152.
  7. Eliel, E.L.; Frye, S.V.; Hortelano, E.R.; Chen, X.; Bai, X. Pure Appl. Chem., 1991, 63, 1591 and refs. cited therein; Nishida, M.; Nakaoka, K.; Ono, S.; Yonemitsu, O.; Nishida, A.; Kawahara, N.; Takayanagi, H. J. Org. Chem., 1993, 58, 5870; Solladie, G; Lohse, O. Tetrahedron: Asymmetry, 1993, 4, 1547; Wei, J.; Hutchins, R.O.; Prol Jr., J. J. Org. Chem., 1993, 58, 2920.
  8. a) Kita, Y.; Shibata, N.; Kawano, N.; Kukui, S.; and Fujimori, C. Tetrahedron Lett., 1994, 35, 3575; b)
- 8. a) Kita, Y.; Shibata, N.; Kawano, N.; Kukui, S.; and Fujimori, C. Tetrahedron Lett., 1994, 35, 3575; b) Kita, Y.; Shibata, N.; Yoshida, N.; Fukui, S.; Fujimori, C. Tetrahedron Lett., 1994, 35, 2569; c) Kita, Y.; Shibata, N.; Yoshida, N. Tetrahedron Lett., 1993, 34, 4063. For an excellent comprehensive review on the Pummerer reaction see: De Lucchi, O.; Miotti, U.; Modena, G. Organic Reactions, 1990, 40, 157.
- 9. Asymmetric synthesis of lamivudine: Milton, J.; Brand, S.; Jones, M.F.; Rayner, C.M.; submitted for publication. A related racemic synthesis using a Pummerer rearrangement as a key step has been carried out, see: Dwyer, O, submitted for publication.
- 10. Thiols were prepared from the corresponding alkyl halides via the xanthate ester and liberation of the thiol using ethylene diamine: Djerassi, C.; Gorman, M.; Markley, F.X.; Oldenberg, E.B. J. Amer. Chem. Soc., 1955,77, 568; Mori, K.; Nakamura, Y.; J. Org. Chem., 1969, 34, 4170. Menthyl refers to esters of (1R,2S,5R)-(-)-menthol. (2R)-(Cbz)Cys(OMe) (entry 16) was prepared according to the literature procedure: Zervas, L.; Photaki, I. J. Amer. Chem. Soc., 1962, 84, 3887.
- 11. Kelly, T.R.; Schmidt, T.E.; Haggerty, J.G. Synthesis, 1972, 544.
- 12. All new compounds were characterised by <sup>1</sup>H and <sup>13</sup>C NMR, IR, and mass spectra, and gave satisfactory elemental analysis and/or accurate mass spectra unless otherwise stated.
- 13. Cygler, M.; Grochulski, P.; Kazlauskas, R.J.; Schrag, J.D.; Bouthillier, F.; Rubin, B.; Serreqi, A.N.; Gupta, A.J. J. Amer. Chem. Soc., 1994, 116, 3180 and refs. cited therein.
- 14. Evans, C.T.; Roberts, S.M.; Shoberu, K.A.; Sutherland, A.G. J. Chem. Soc., Perkin Trans. 1, 1992, 589. See also ref. 1a, section 1:10.1.